

# Single Cell and Single Molecule Analysis of Cancer

Michael Schatz

April 20, 2015  
Laufer Center Retreat



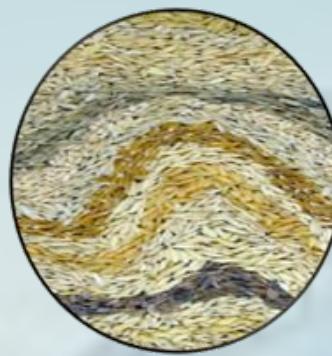
# Schatzlab Overview



## Human Genetics

Role of mutations in disease

Narzisi *et al.* (2014)  
Iossifov *et al.* (2014)



## Plant Biology

Genomes & Transcriptomes

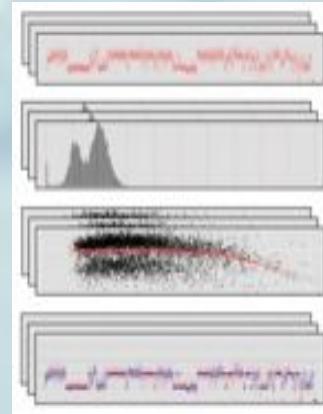
Schatz *et al.* (2014)  
Ming *et al.* (2013)



## Algorithmics & Systems Research

Ultra-large scale biocomputing

Marcus *et al.* (2014)  
Schatz *et al.* (2013)



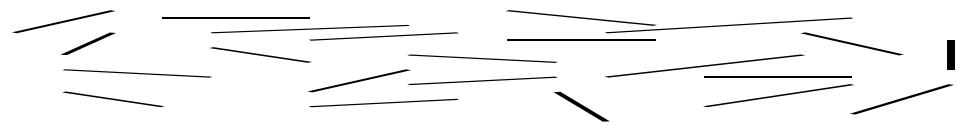
## Single Cell & Single Molecule

CNVs, SVs, & Cell Phylogenetics

Garvin *et al.* (2014)  
Roberts *et al.* (2013)

# Sequence Assembly Problem

## 1. Shear & Sequence DNA



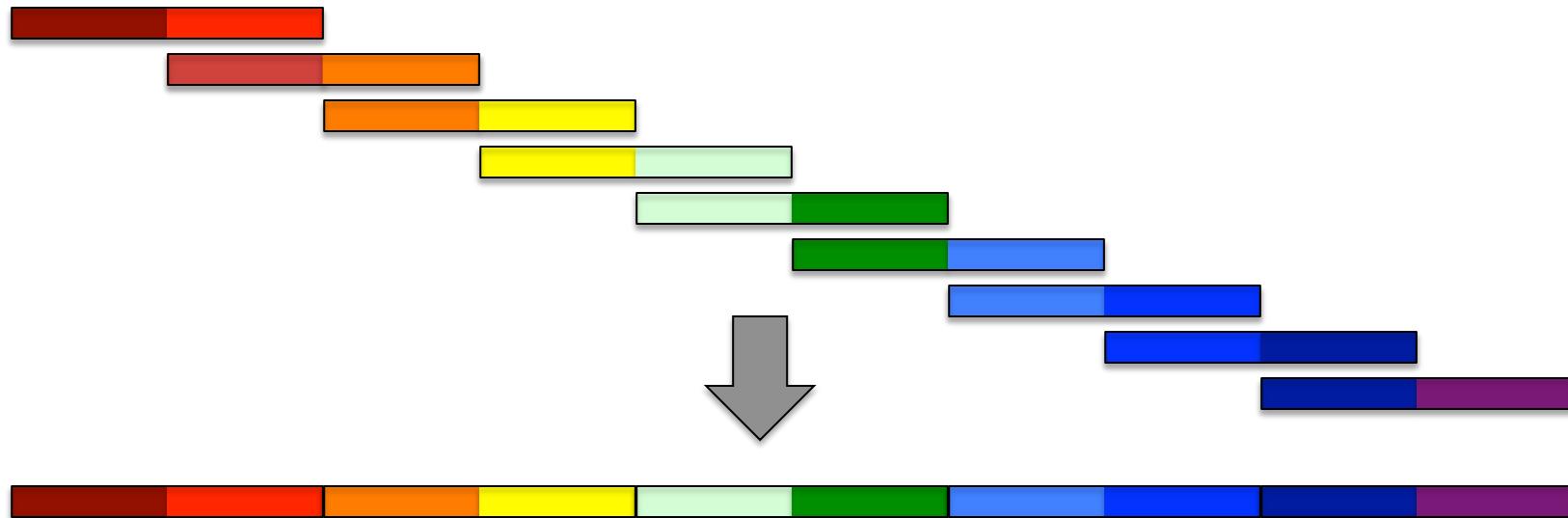
## 2. Construct assembly graph from overlapping reads

...AGCCTAGGGATGCGCGACACGT

GGATGCGCGACACGT CGCATATCCGGTTGGT CAACCTCGGACGGAC

CAACCTCGGACGGAC CTCAGCGAA...

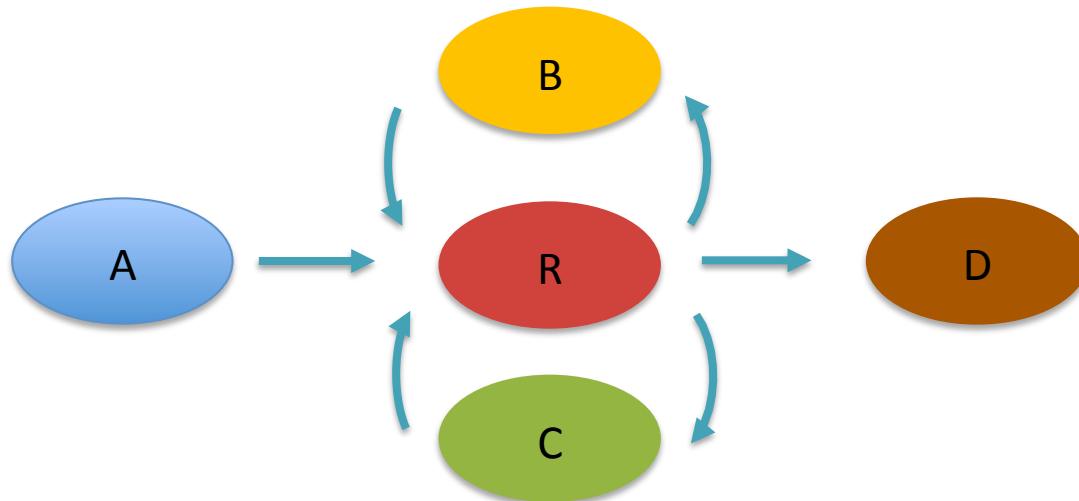
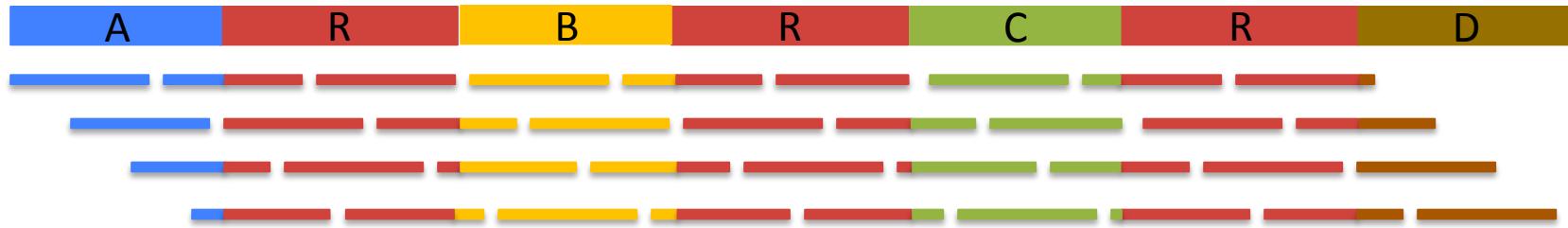
## 3. Simplify assembly graph



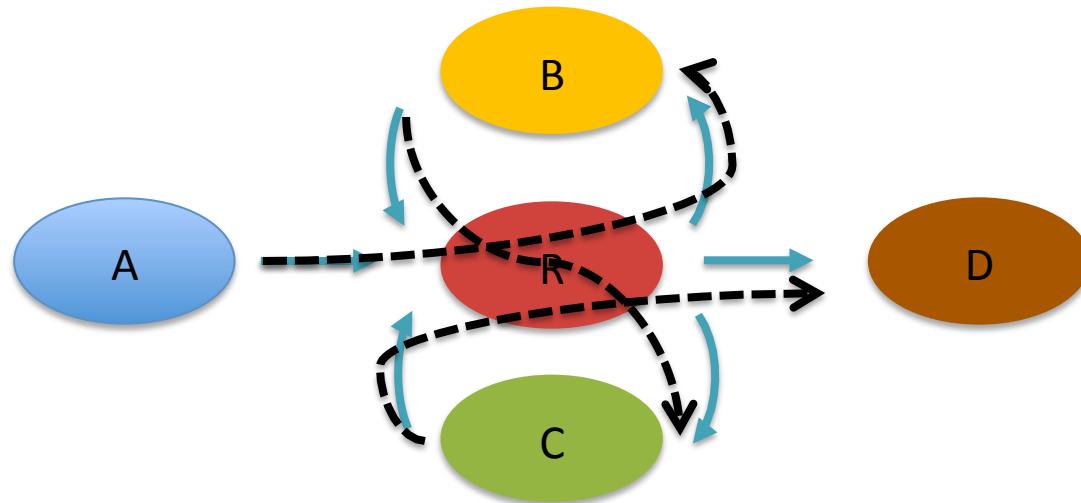
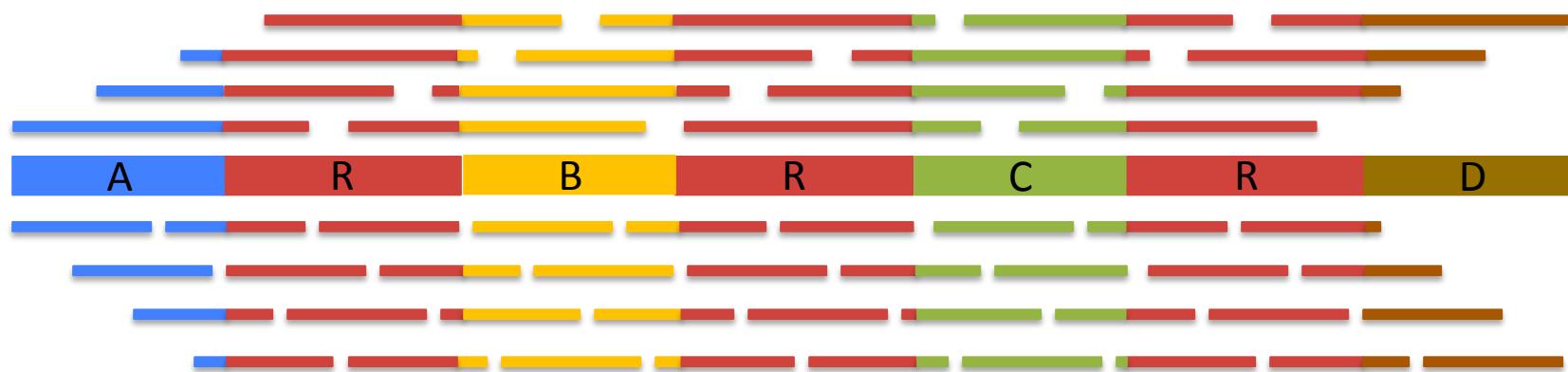
### On Algorithmic Complexity of Biomolecular Sequence Assembly Problem

Narzisi, G, Mishra, B, Schatz, MC (2014) *Algorithms for Computational Biology*. Lecture Notes in Computer Science. Vol. 8542

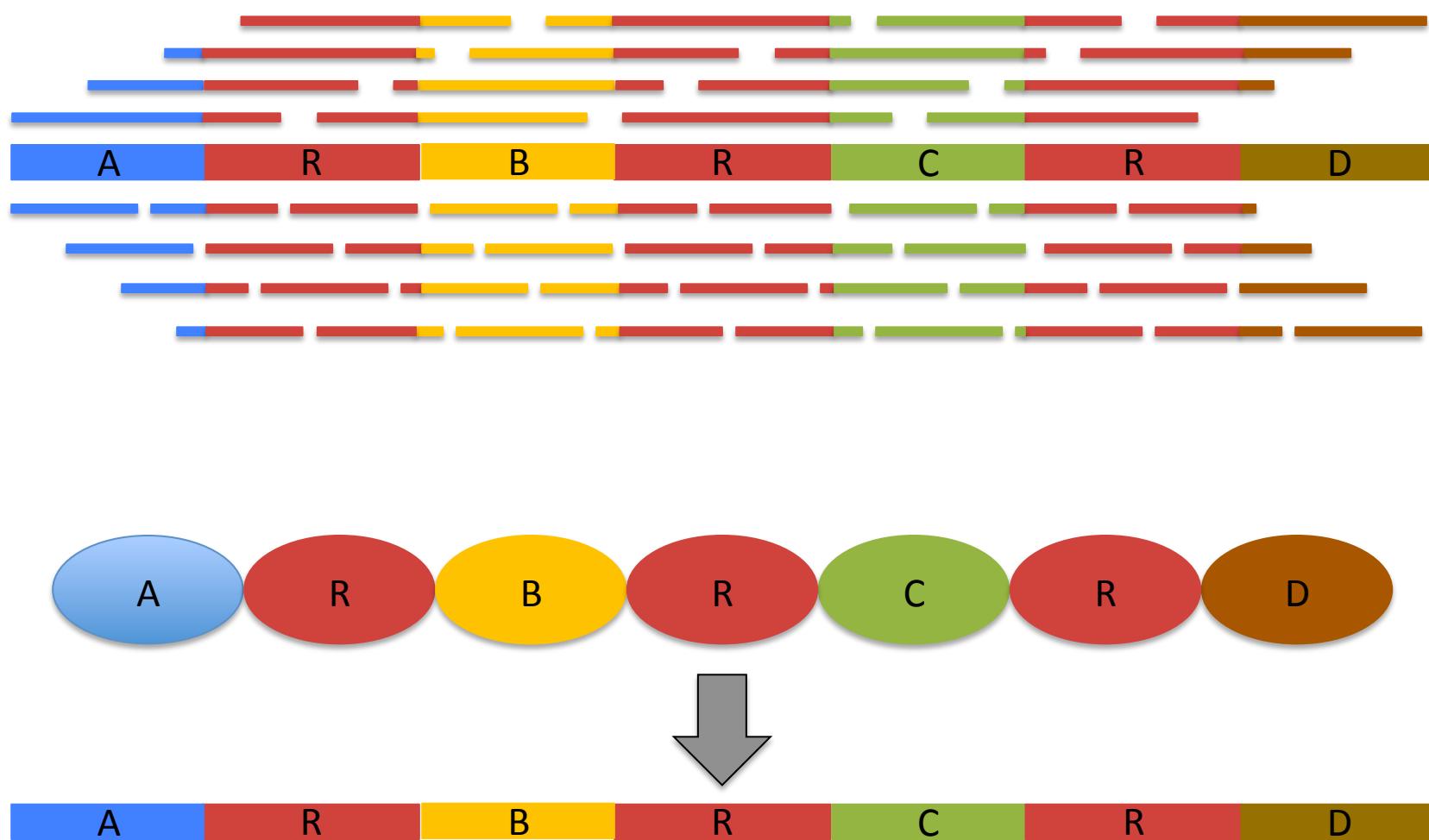
# Assembly Complexity



# Assembly Complexity



# Assembly Complexity



**The advantages of SMRT sequencing**

Roberts, RJ, Carneiro, MO, Schatz, MC (2013) *Genome Biology*. 14:405

# Genomics Arsenal in the year 2015

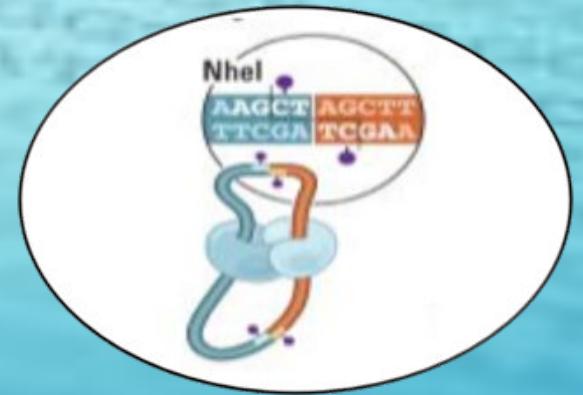
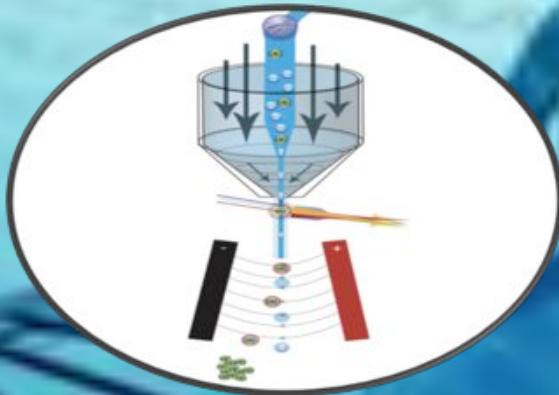
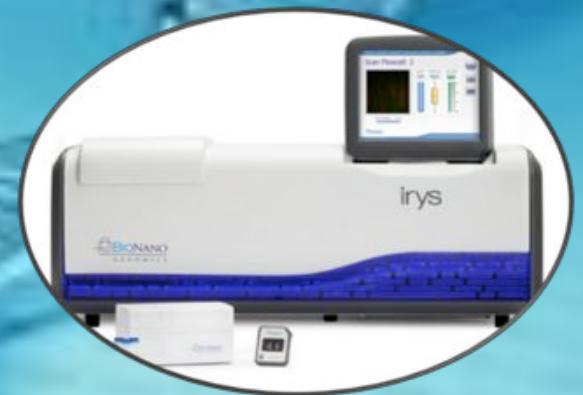
Sample Preparation



Sequencing

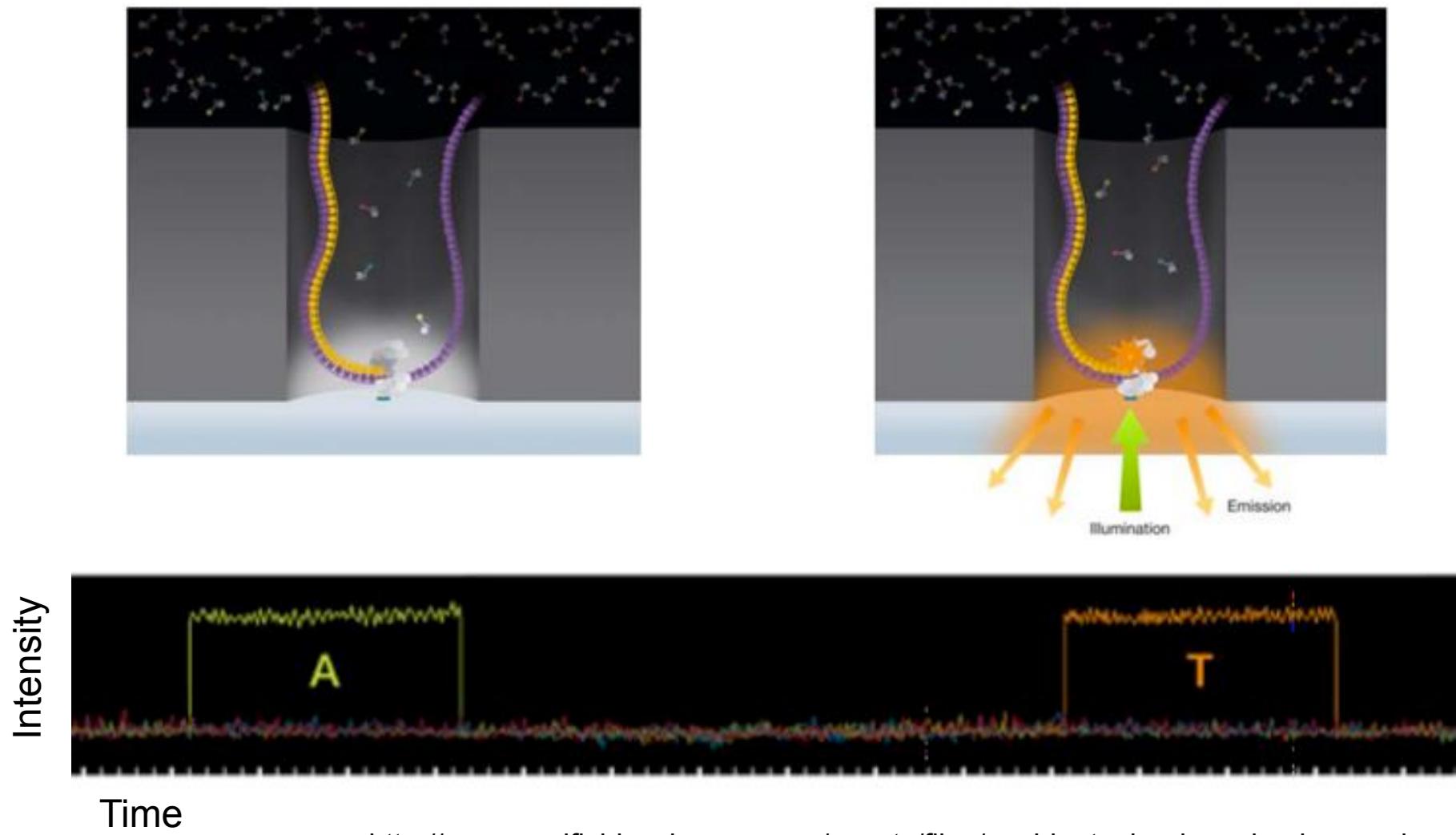


Chromosome Mapping

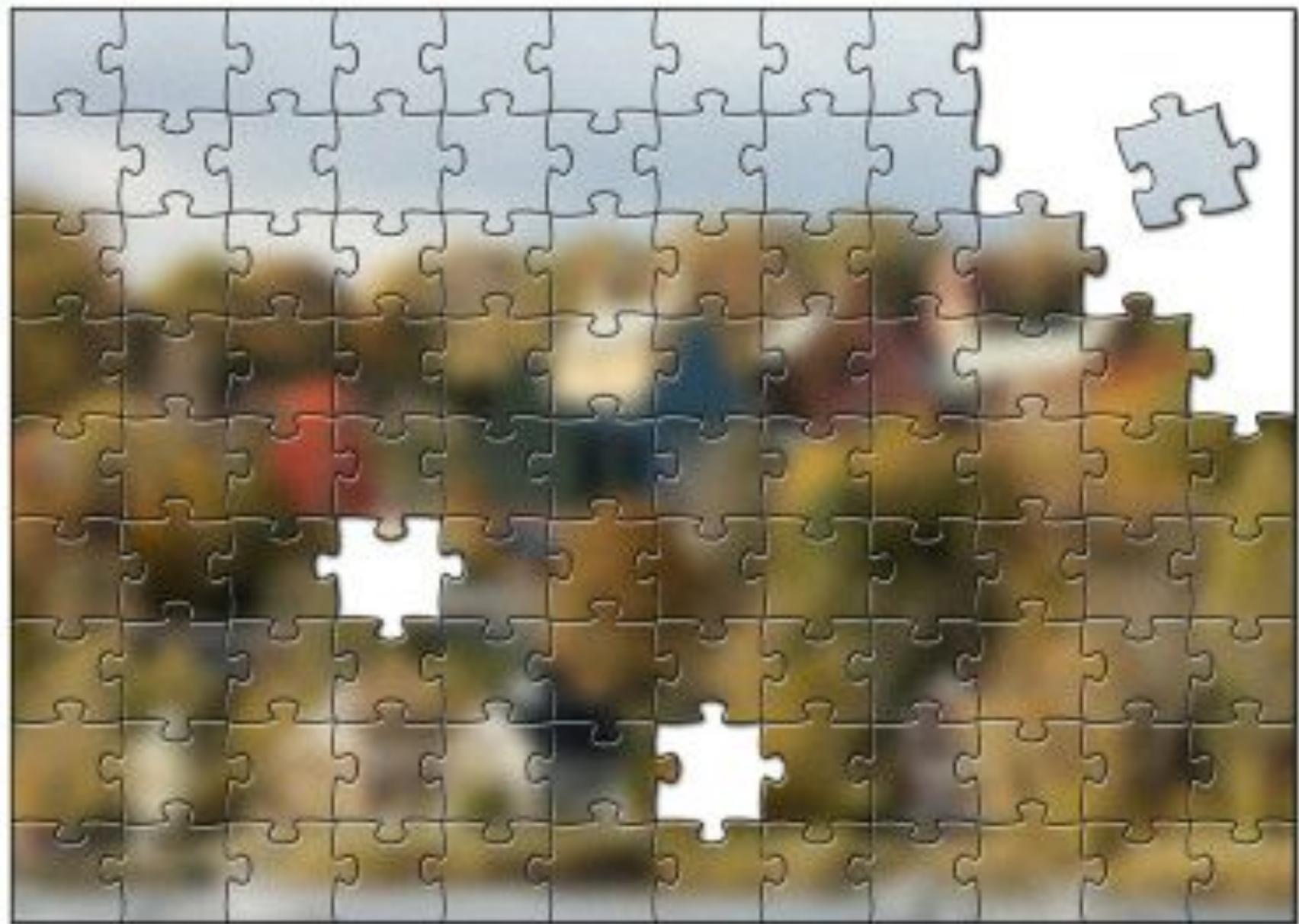


# PacBio SMRT Sequencing

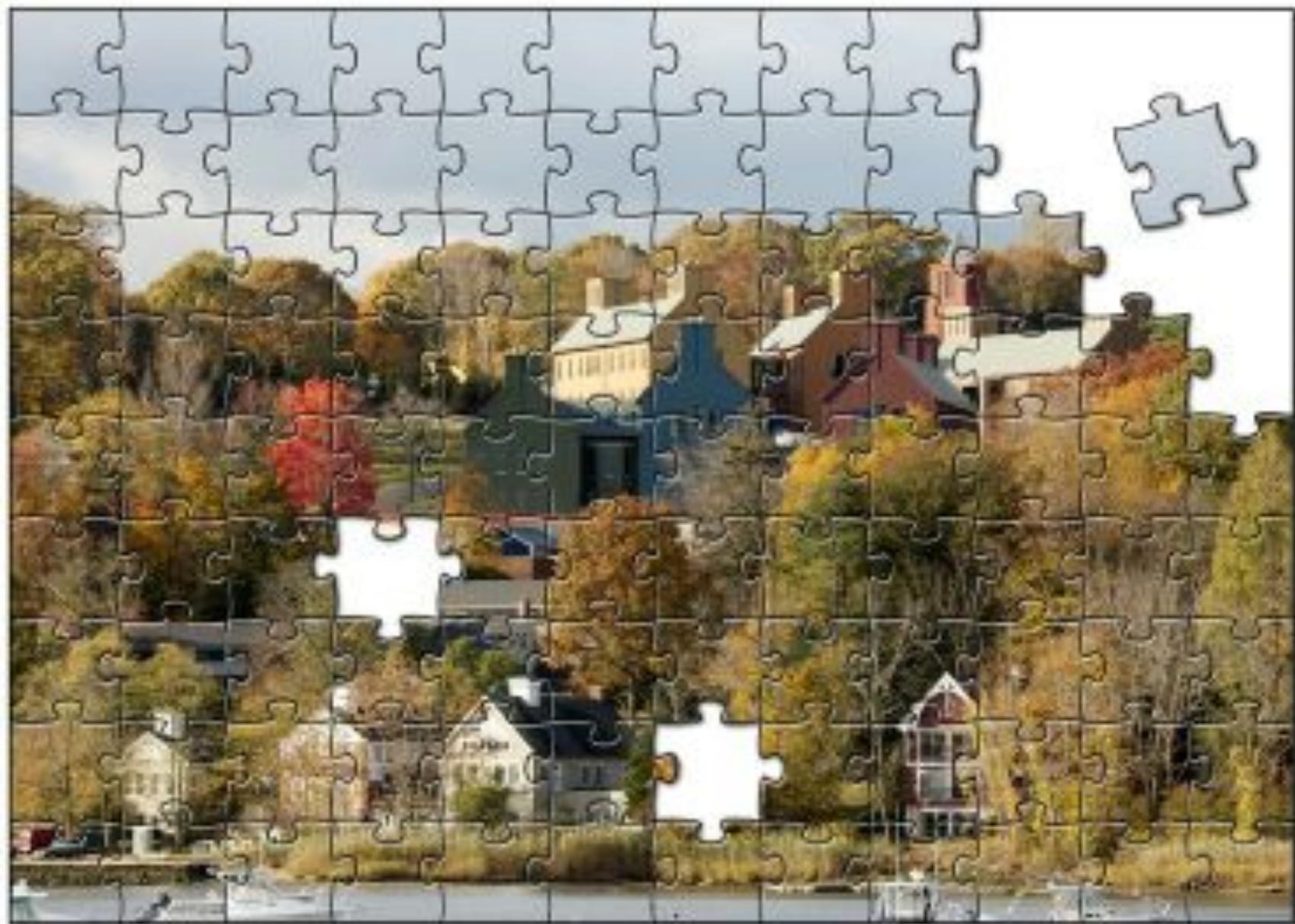
Imaging of fluorescently phospholinked labeled nucleotides as they are incorporated by a polymerase anchored to a Zero-Mode Waveguide (ZMW).



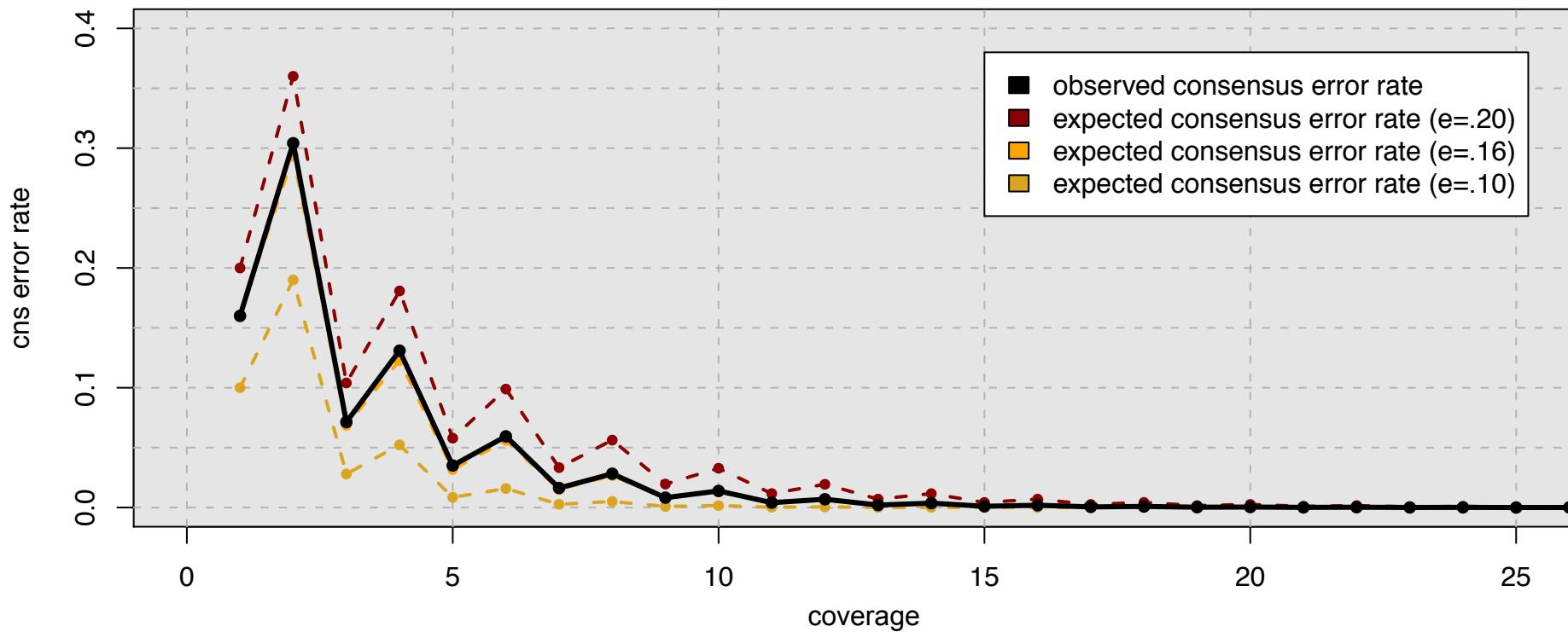
# Single Molecule Sequences



# “Corrective Lens” for Sequencing



# Consensus Accuracy and Coverage



Coverage can overcome random errors

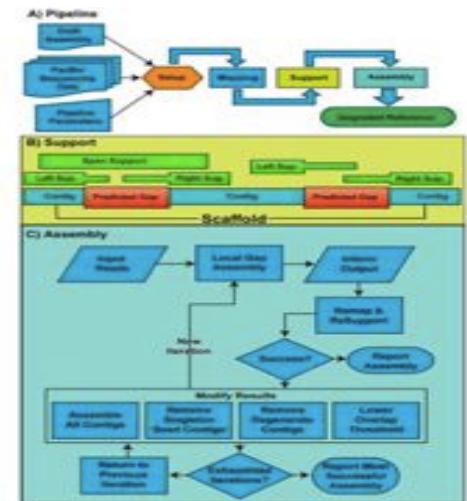
- Dashed: error model from binomial sampling
- Solid: observed accuracy

Koren, Schatz, et al (2012)  
Nature Biotechnology. 30:693–700

$$CNS\ Error = \sum_{i=\lceil c/2 \rceil}^c \binom{c}{i} (e)^i (1-e)^{n-i}$$

# PacBio Assembly Algorithms

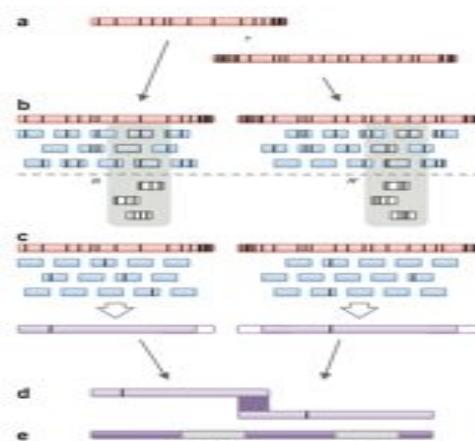
## PBJelly



### Gap Filling and Assembly Upgrade

English et al (2012)  
PLOS One. 7(11): e47768

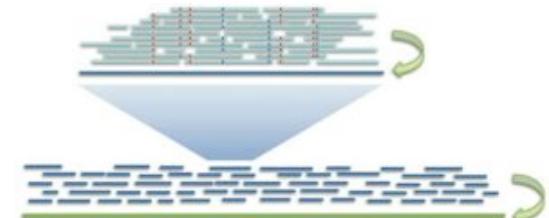
## PacBioToCA & ECTools



### Hybrid/PB-only Error Correction

Koren, Schatz, et al (2012)  
Nature Biotechnology. 30:693–700

## HGAP & Quiver



$$\Pr(\mathbf{R} \mid T)$$
$$\Pr(\mathbf{R} \mid T) = \prod_k \Pr(R_k \mid T)$$

Quiver Performance Results Comparison to Reference Genome ( <i>M. ruber</i> ; 3.1 MB; SMRT® Cells)		
	Initial Assembly	Quiver Consensus
QV	43.4	54.5
Accuracy	99.99540%	99.99964%
Differences	141	11

### PB-only Correction & Polishing

Chin et al (2013)  
Nature Methods. 10:563–569

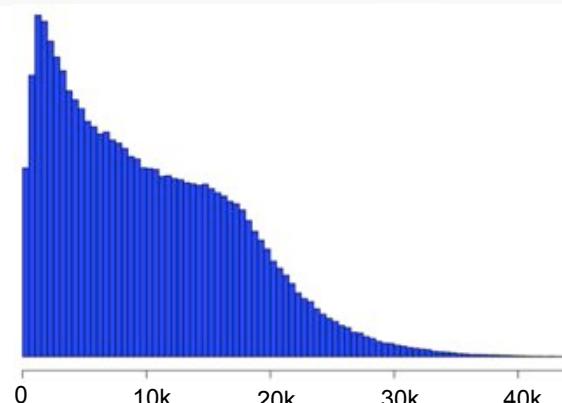
< 5x

PacBio Coverage

> 50x

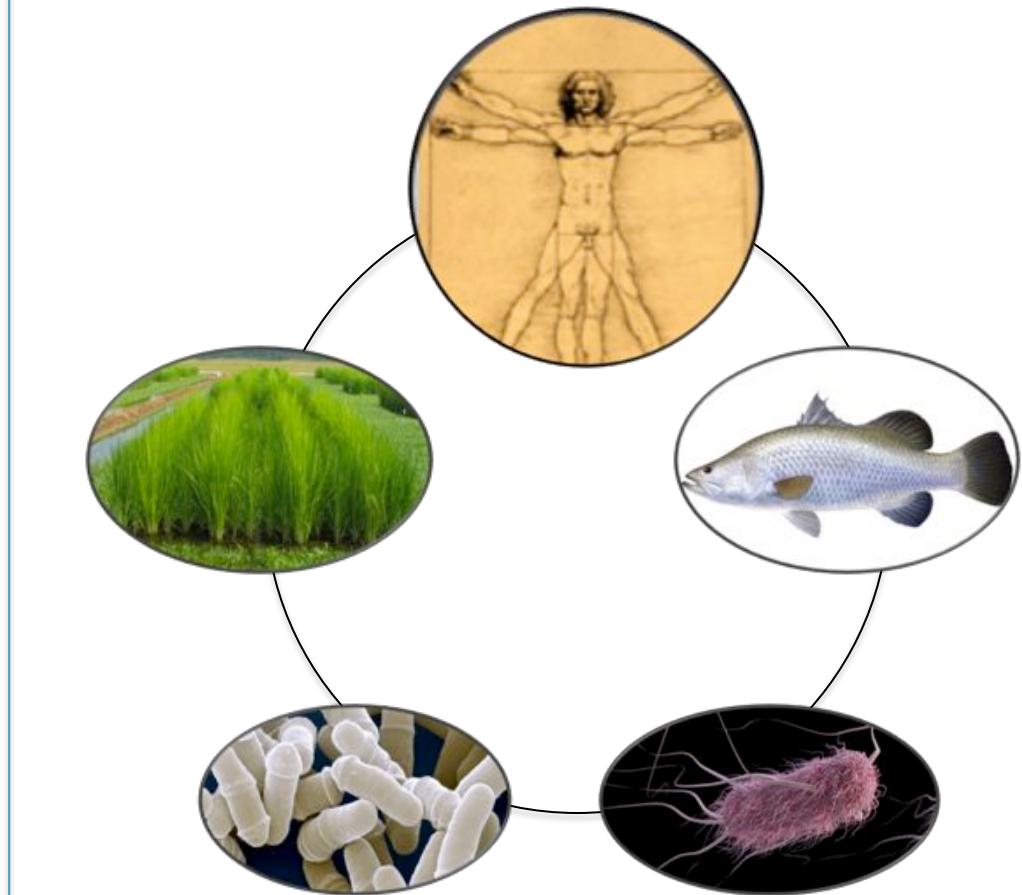
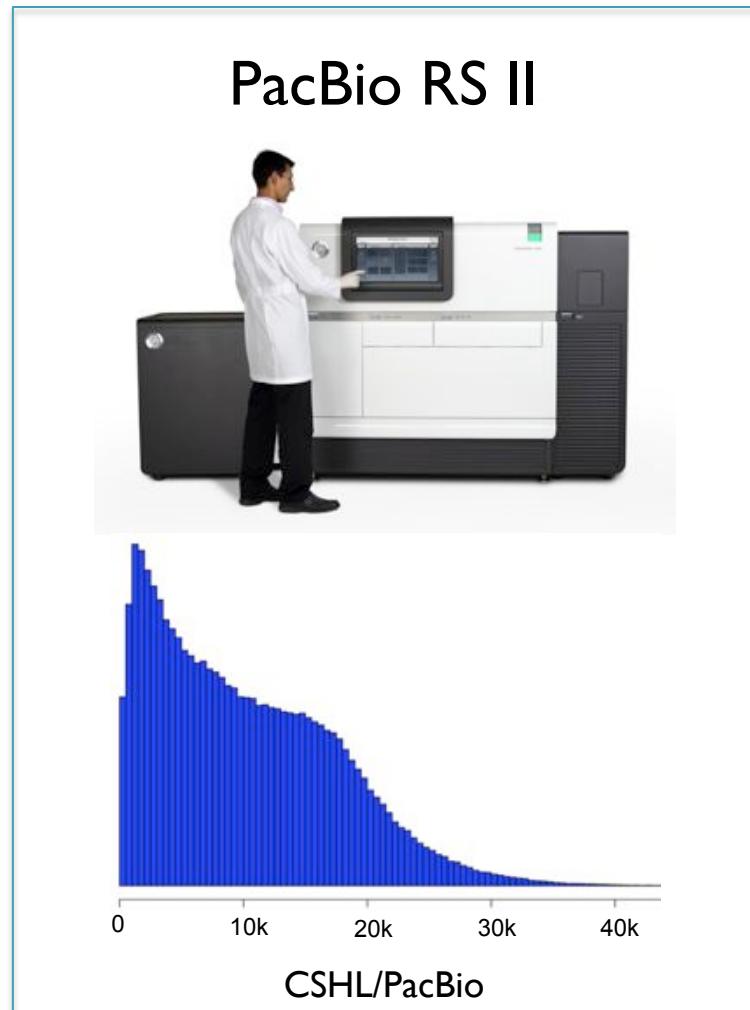
# 3<sup>rd</sup> Gen Long Read Sequencing

PacBio RS II

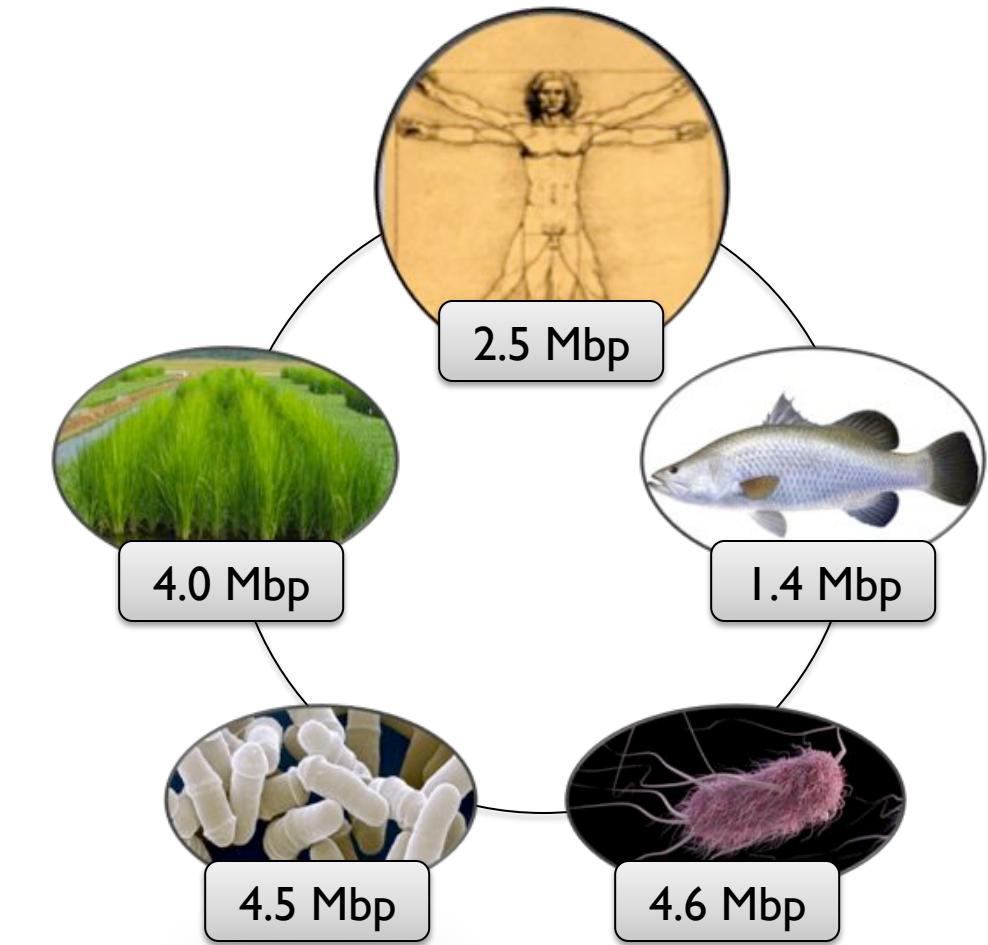
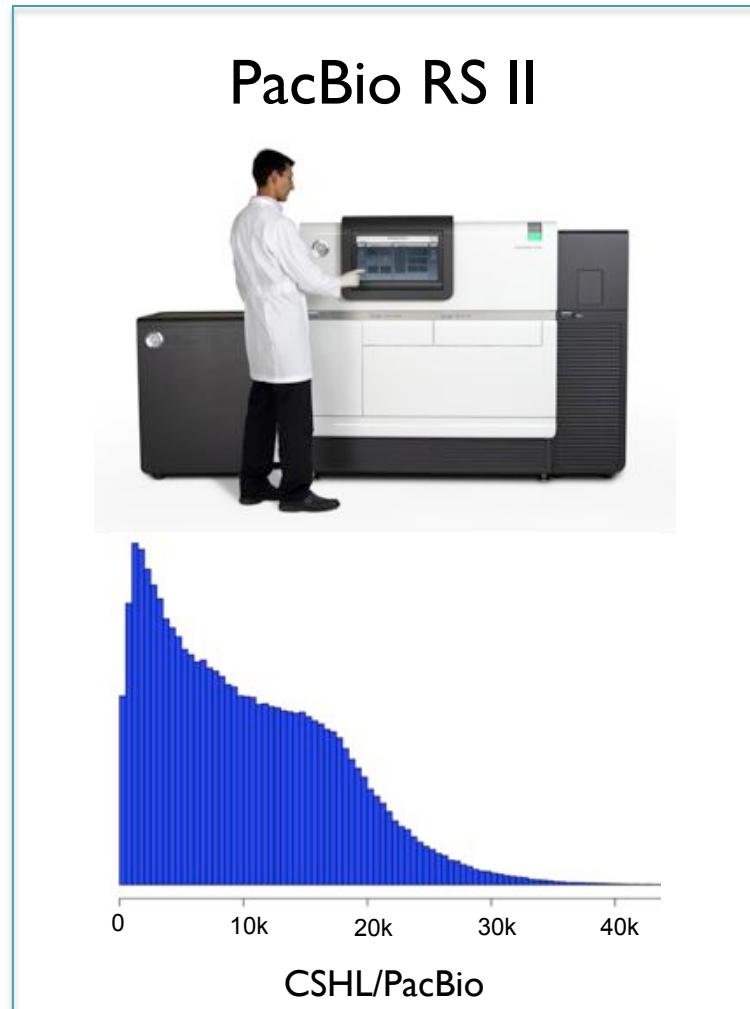


CSHL/PacBio

# 3<sup>rd</sup> Gen Long Read Sequencing



# 3<sup>rd</sup> Gen Long Read Sequencing



# SK-BR-3



Most commonly used Her2-amplified breast cancer cell line

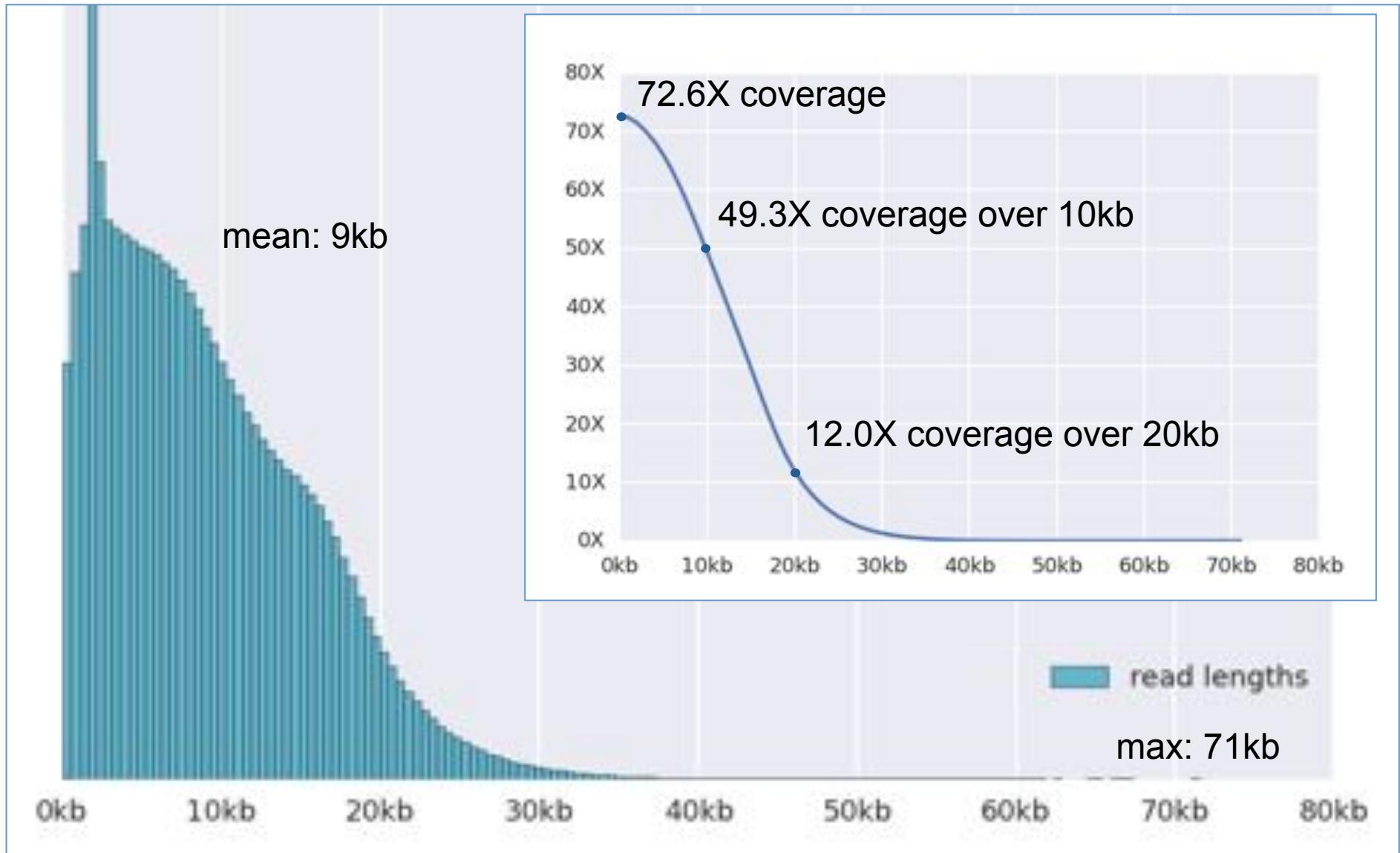


(Davidson et al, 2000)

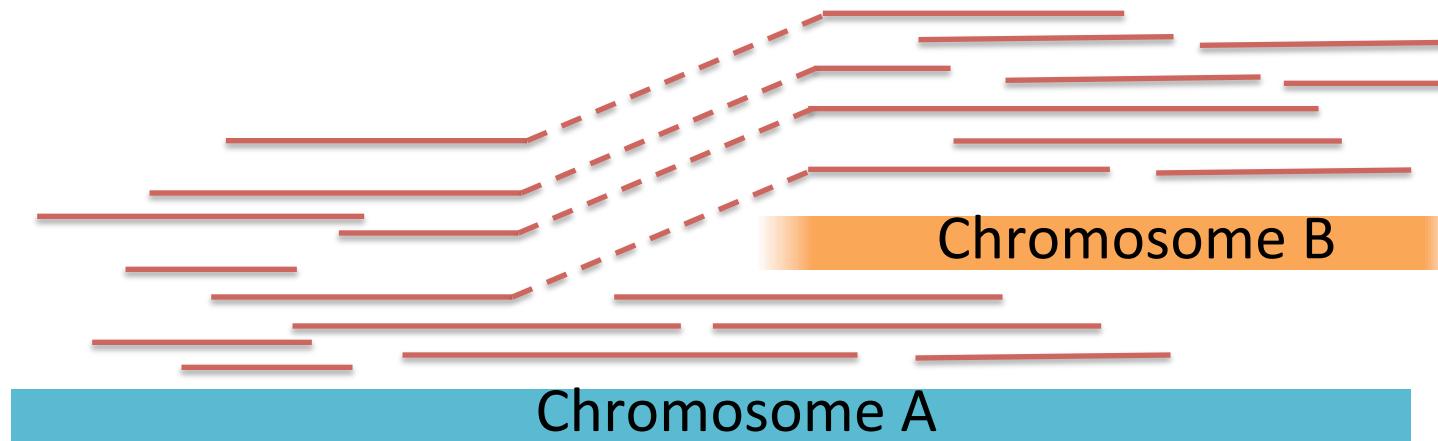
***Can we resolve the complex structural variations, especially around Her2?***

Ongoing collaboration between CSHL and OICR to *de novo* assemble  
the complete cell line genome with PacBio long reads

# PacBio read length distribution



# Structural variant discovery with long reads



## 1. Alignment-based split read analysis: Efficient capture of most events

BWA-MEM + Lumpy

## 2. Local assembly of regions of interest: In-depth analysis with *base-pair precision*

Localized HGAP + Celera Assembler + MUMmer

## 3. Whole genome assembly: In-depth analysis including *novel sequences*

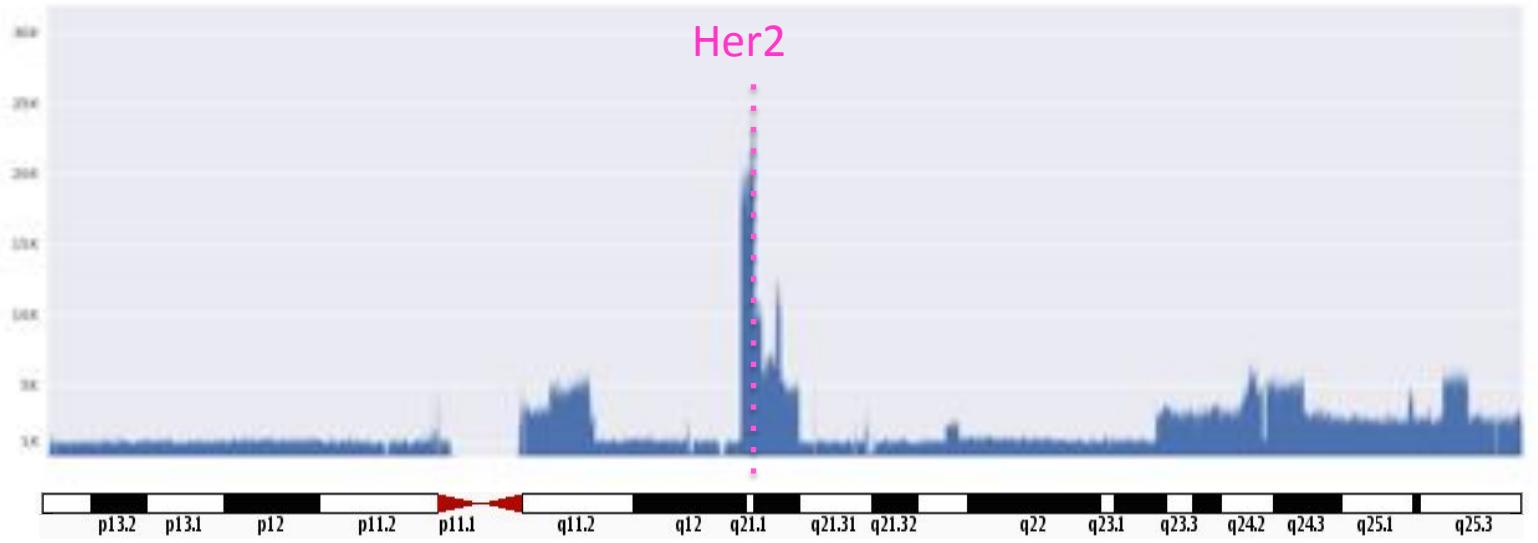
DNAexus-enabled version of Falcon

Total Assembly: 2.64Gbp

Contig N50: 2.56 Mbp

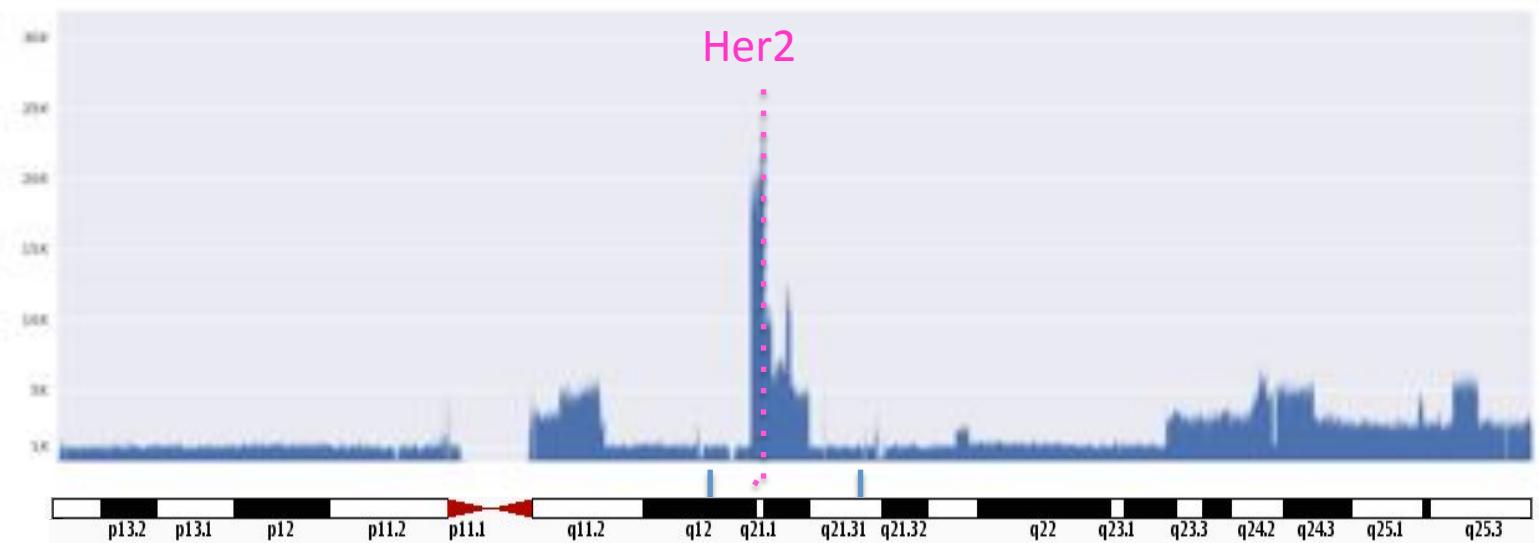
Max Contig: 23.5Mbp

PacBio



Chr 17: 83 Mb

PacBio



PacBio  
chr17

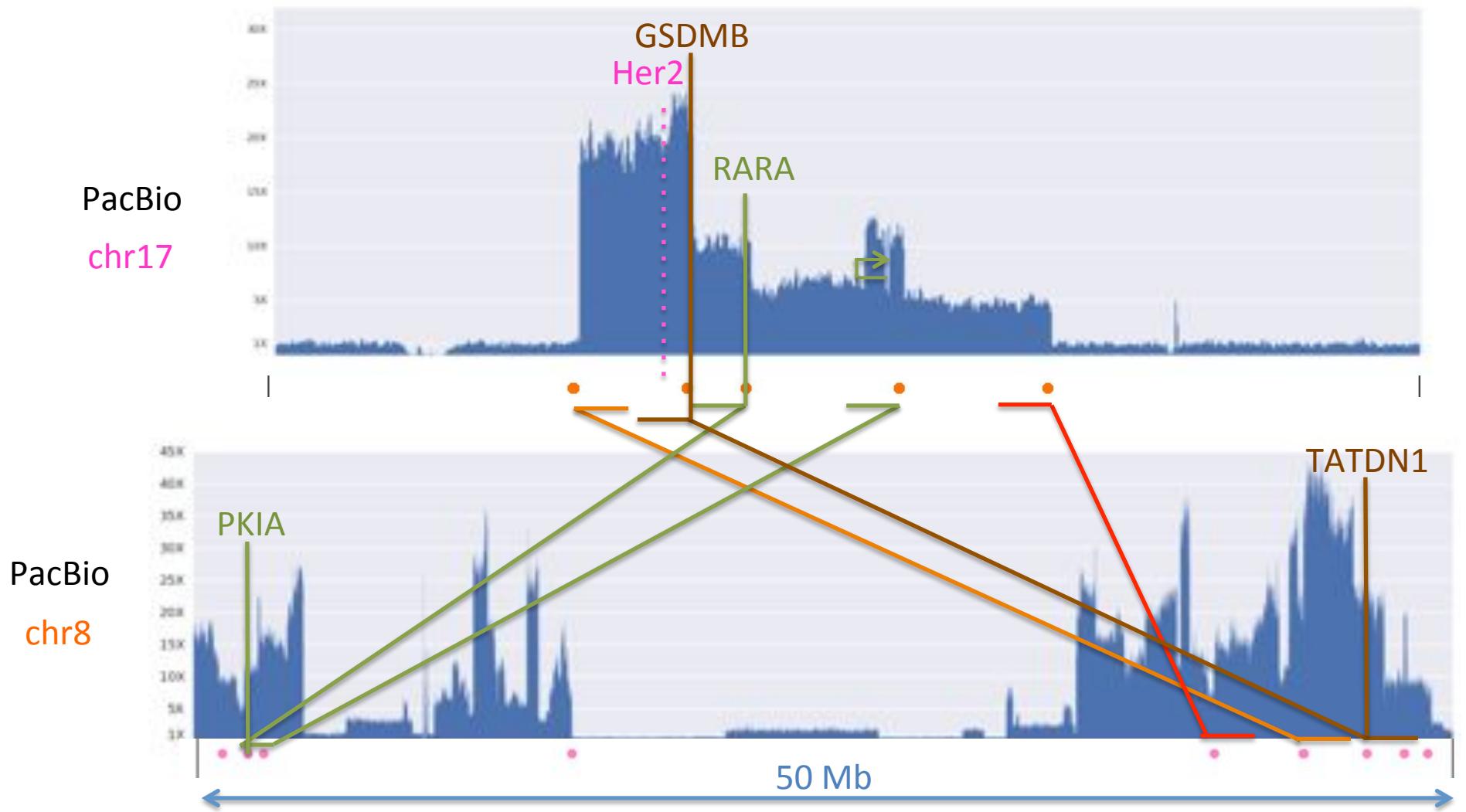


PacBio

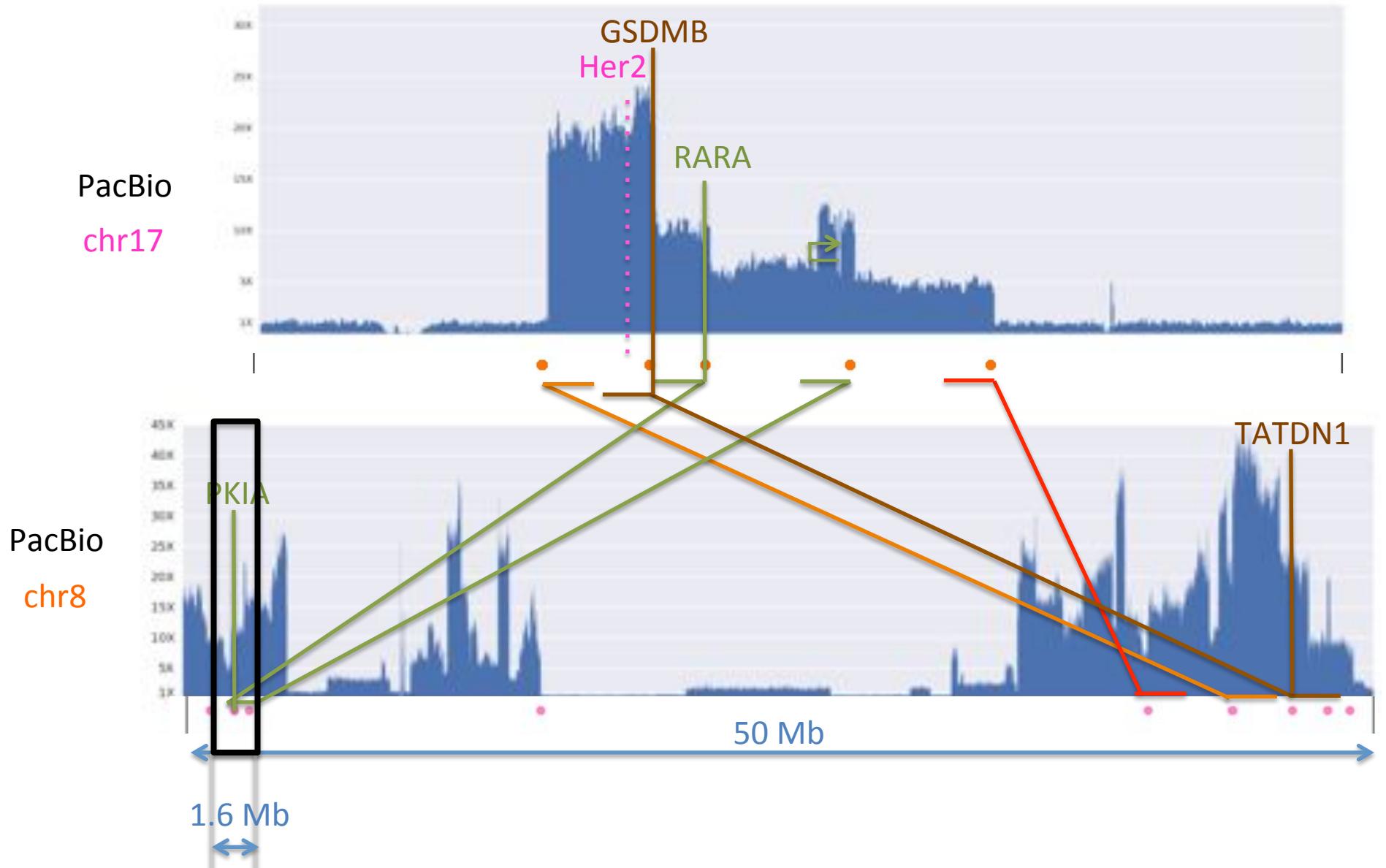


PacBio  
chr17

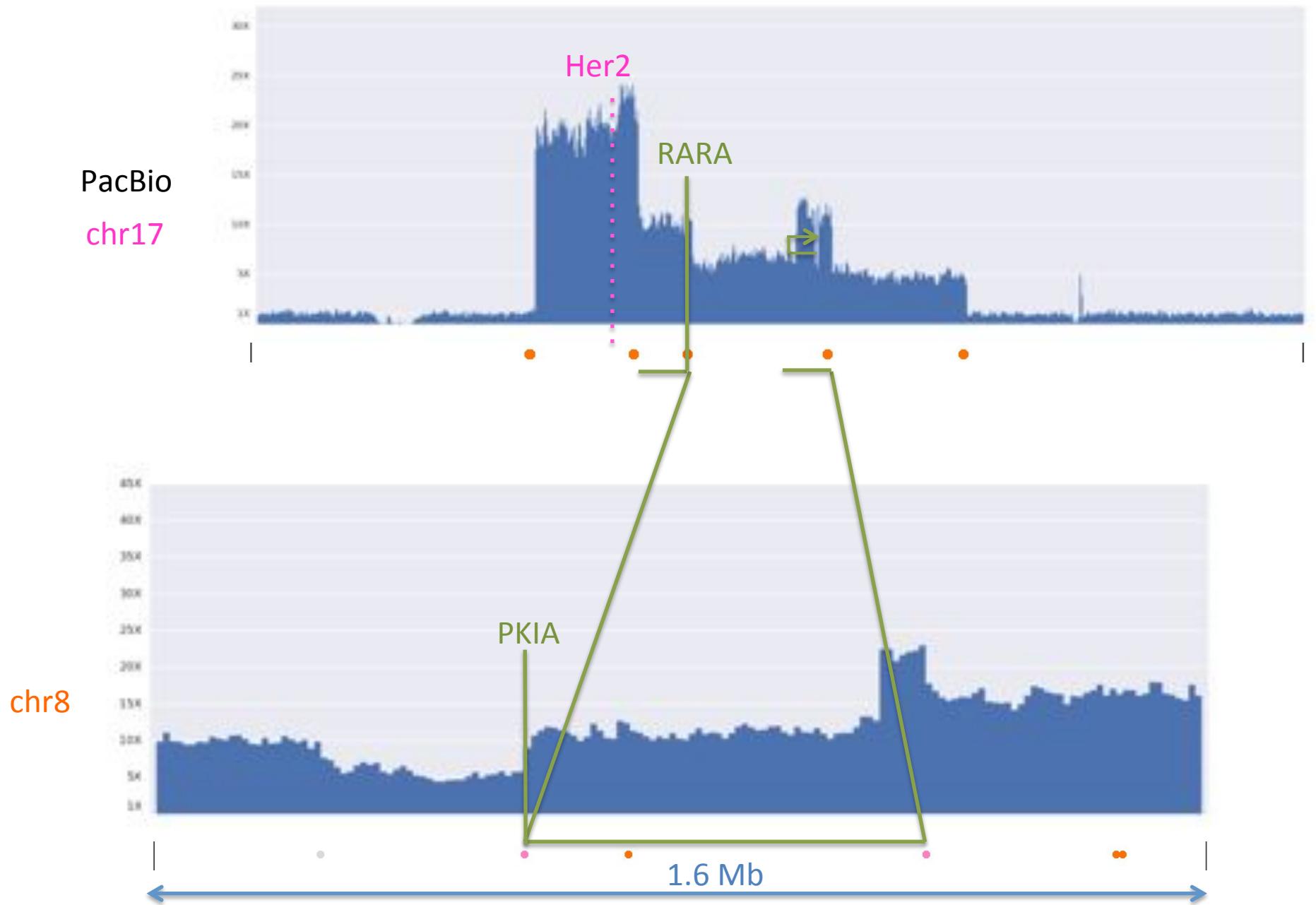




Confirmed both known gene fusions in this region



Confirmed both known gene fusions in this region



Joint coverage and breakpoint analysis to discover underlying events

# Cancer lesion Reconstruction



By comparing the proportion of reads that are spanning or split at breakpoints we can begin to infer the history of the genetic lesions.

1. Healthy diploid genome
2. Original translocation into chromosome 8
3. Duplication, inversion, and inverted duplication within chromosome 8
4. Final duplication from within chromosome 8

# Cancer lesion Reconstruction

**Available *today* under the Toronto Agreement:**

- Fastq & BAM files of aligned reads
- Interactive Coverage Analysis with BAM.IOBIO
- Whole genome assembly & alignment

**Available soon**

- Whole genome methylation analysis
- Full length cDNA transcriptome analysis
- Comparison to single cell analysis of >100 individual cells

***Go see Maria's poster!***

4. Final duplication from within chromosome 8

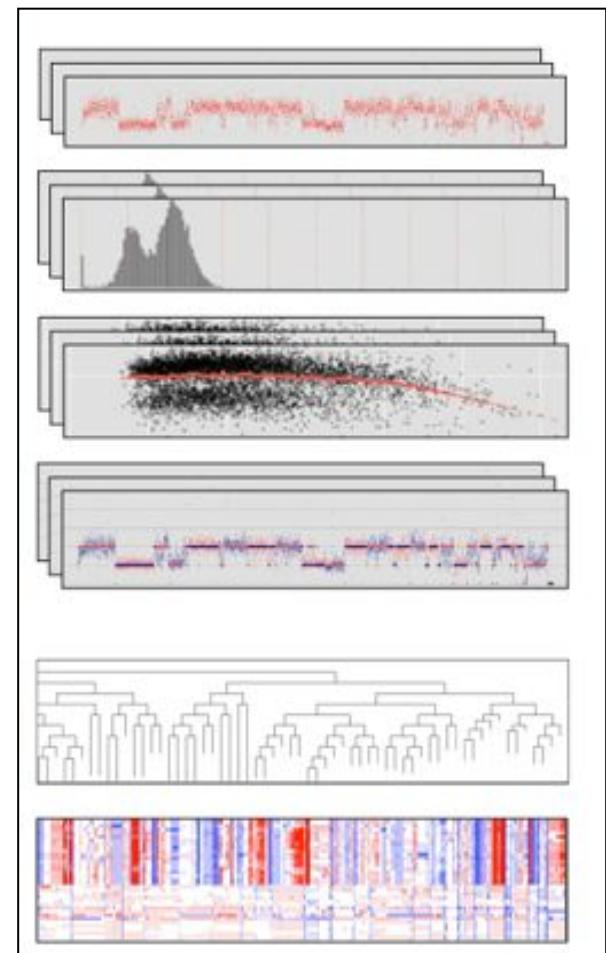
# Gingko

<http://qb.cshl.edu/ginkgo>



## **Single-Cell Copy Number Analysis**

- Extremely low coverage sequencing ( $\sim .1x$ ) from amplified cells is sufficient to determine large copy number changes (>50kbp)
- Use this technique to discover CNVs in multiple cells from the same tumor to map its progress
- Implemented a new analysis suite (Ginkgo) to carry out the highly specialized processing



**Interactive analysis and quality assessment of single-cell copy-number variations.**

Garvin, T., Aboukhalil, et al. (2015) Under review

# What should we expect from an assembly?

## ***Summary & Recommendations***



L

New Results

**Error correction and assembly complexity of single molecule sequencing reads.**

Hayan Lee , James Gurtowski , Shinjae Yoo , Shoshana Marcus , W. Richard McCombie , Michael Schatz  
doi: <http://dx.doi.org/10.1101/006395>

> 5GB: Email [mschatz@cshl.edu](mailto:mschatz@cshl.edu)

V

C

***The year 2015 will mark the return to reference quality genome sequence***

Technologies are quickly improving, exciting new sequencing technologies

# Acknowledgements

## **Schatz Lab**

Rahul Amin  
Eric Biggers  
Han Fang  
Tyler Gavin  
James Gurtowski  
Ke Jiang  
Hayan Lee  
Zak Lemmon  
Shoshana Marcus  
Giuseppe Narzisi  
Maria Nattestad  
Aspyn Palatnick  
Srividya  
Ramakrishnan  
Fritz Sedlazeck  
Rachel Sherman  
Greg Verture  
Alejandro Wences

## **CSHL**

Hannon Lab  
Gingeras Lab  
Jackson Lab  
Hicks Lab  
Iossifov Lab  
Levy Lab  
Lippman Lab  
Lyon Lab  
Martienssen Lab  
McCombie Lab  
Tuveson Lab  
Ware Lab  
Wigler Lab

## **SBU**

Skiena Lab  
Patro Lab

## **Cornell**

Susan McCouch  
Lyza Maron  
Mark Wright

## **OICR**

John McPherson  
Karen Ng  
Timothy Beck  
Yogi Sundaravadanam

## **NBACC**

Adam Phillippy  
Serge Koren



# **Genome Informatics**

Janet Kelso, Daniel MacArthur, Michael Schatz

Oct 28 - 31, 2015



# Thank you

<http://schatzlab.cshl.edu>

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